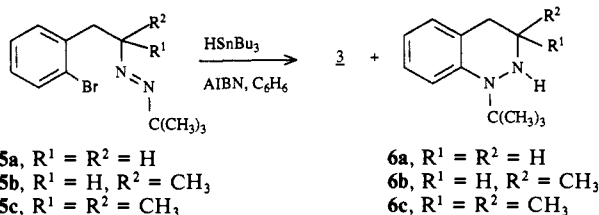
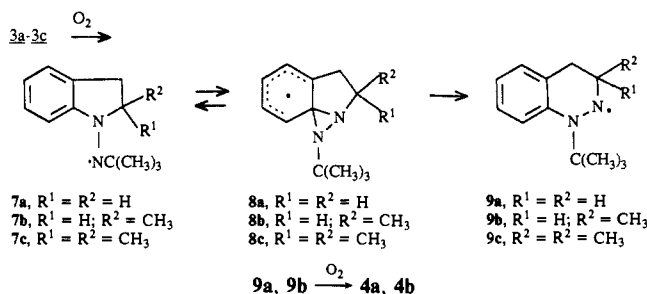


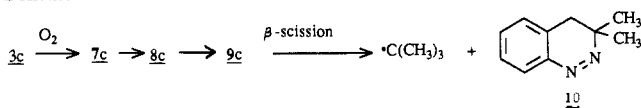
Scheme I



Scheme II



Scheme III



hydrazyl (9) readily accounts for the observations.

The mechanism of Scheme II is supported strongly by the behavior of 3c, which afforded 3,4-dihydrocinnoline (10), Scheme III. Hydrazyl 9c would be expected to lose the *tert*-butyl radical, rather than the CH₃ radical, by β -scission. There is precedent for facile loss of *tert*-butyl from a hydrazyl¹⁰ except in cases of fixed and unfavorable geometry.¹¹

It was possible to observe hydrazyl 7c by irradiation of a solution of 3c at -40 °C in a mixed solvent consisting of di-*tert*-butyl peroxide and CH₂Cl₂ (1:1 by volume), in the cavity of an EPR spectrometer. Although the initial spectrum was complex, probably because of an impurity present, continued irradiation led quickly to a steady five-line spectrum (intensity ratios within 20% of 1:2:3:2:1 at $g = 2.0038$). The g value is close to those of typical hydrazyls,¹² and the observation of five-lines as described implies that $a_{N(1)} \approx a_{N(2)} \approx 11.8$ G. In most hydrazyls,¹² $a_{N(1)} > a_{N(2)}$ but the reverse is known,¹³ and it is therefore not surprising that $a_{N(1)} \approx a_{N(2)}$ should be encountered, as is the case for 7c. The signal faded rapidly when the light was turned off.

Increasing the temperature did not lead to a change in the spectrum, indicating either that neophyl rearrangement is slow or that the rearranged radical 9c loses the *tert*-butyl group very rapidly. A strong indication that the latter is true came from attempts to prepare 9c directly from 6c, by the method described above for the generation of 7c from 3c. No signal at all was observable. Since it is highly unlikely that *tert*-butoxyl radicals abstract efficiently from 3c but not from 6c, the conclusion is that loss of *tert*-butyl from 9c is very much faster than the neophyl rearrangement of 7c to 9c.

The sequence of reactions of 3, induced by oxygen, represents the first examples of neophyl rearrangements of hydrazyls to isomeric hydrazyls. Rearrangement of radicals 7 is surprising because hydrazyls are normally stabilized by three-electron π -bonding. Recent results, that add to an extensive body of literature on the subject of bonding in hydrazyls, include the observation that the NH₂ hydrogens of 1-phenylhydrazyl are nonequivalent¹⁴ and calculations of torsional and inversion barriers for hydrazyl

itself.¹⁵ In the case of radicals 7, hydrazyl resonance is probably less important because it requires the *tert*-butyl group to be held, with entropic cost, in or near the molecular plane, where it must interact sterically with either the CR¹R² moiety or with the "peri" hydrogen at C-7. Although radicals 9 also have a "peri" interaction of the type mentioned above, hydrazyl resonance comes at lower entropic cost because both N atoms are in a ring. Rearrangement of 7 to 9 can therefore be expected to be exothermic and effectively irreversible.

Neophyl rearrangement could conceivably be involved during the synthesis of 3 and 6 (Scheme I). Beckwith and co-workers¹⁶ have found that 5-exo cyclizations of alkenylaryl and (alkenyl-oxy)aryl radicals are followed by neophyl rearrangements that can compete with H-abstraction from HSnBu₃. In the present cases, the ratios (3:6) were independent of the initial concentration of HSnBu₃ in the range 0.20–1.15 M. That result suggests that the neophyl rearrangements of 7 are relatively slow, as expected,¹⁷ for it is unlikely that the neopentyl-like 7 abstracts from HSnBu₃ with rate constants much larger than those of the Beckwith¹⁶ radicals.

The oxidative rearrangements of 3a and 3b reported here may represent an attractive route to 1-alkyl-1,4-dihydrocinnolines, such as 4a and 4b, from 1-(alkylamino)indolines.

Registry No. 3a, 116302-23-9; 3b, 109638-02-0; 3c, 109637-95-8; 4a, 116302-24-0; 4b, 116302-25-1; 7c, 116302-26-2; 10, 116302-27-3.

(15) Raban, M.; Aviram, K.; Kost, D. *Tetrahedron Lett.* **1985**, 26, 3591.

(16) Abeywickrema, A. N.; Beckwith, A. L. J.; Gerba, S. *J. Org. Chem.* **1987**, 52, 4072.

(17) The rate constant for rearrangement of the neophyl radical is 7.6×10^2 s⁻¹ at 25 °C.¹⁸

(18) Franz, J. A.; Barrows, R. D.; Camaioni, D. M. *J. Am. Chem. Soc.* **1984**, 106, 3964.

Control of Enzyme Enantioselectivity by the Reaction Medium

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Received June 14, 1988

Stereoselectivity is one of the hallmarks of enzymatic catalysis.^{1,2} In principle, enzyme stereoselectivity could be altered by protein engineering³ which would be of profound significance for both enzyme-catalyzed preparative synthesis⁴ and mechanistic biochemistry. Recently, we have observed that upon a transition from water to organic solvents as the reaction medium the enantioselectivity of the protease subtilisin Carlsberg in the reaction of peptide synthesis dramatically relaxes.⁵ If general, this phe-

(1) (a) Fersht, A. *Enzyme Structure and Mechanism*, 2nd ed.; Freeman: New York, 1985; Chapter 8. (b) Walsh, C. *Enzymatic Reaction Mechanisms*; Freeman: San Francisco, 1979; Chapter 2. (c) Jones, J. B.; Beck, J. F. In *Applications of Biochemical Systems in Organic Chemistry*; Jones, J. B., Perlman, D., Sih, C. J., Eds.; Wiley: New York, 1976; Part I, pp 107–401. (d) Retey, J.; Robinson, J. A. *Stereospecificity in Organic Chemistry and Enzymology*; Verlag Chemie: Weinheim, 1982.

(2) Bergmann, M.; Fruton, J. S. *J. Biol. Chem.* **1937**, 117, 189–202. Hein, G. E.; Niemann, C. *J. Am. Chem. Soc.* **1962**, 84, 4495–4503. Cohen, S. G.; Milovanovic, A.; Schultz, R. M.; Weinstein, S. Y. *J. Biol. Chem.* **1969**, 244, 2664–2674. Blow, D. M. *The Enzymes* **1971**, 3, 185–212.

(3) Fersht, A. R.; Shi, J.; Wilkinson, A. J.; Blow, D. M.; Carter, P.; Waye, M. M. Y.; Winter, G. P. *Angew. Chem., Int. Ed. Engl.* **1984**, 23, 467–473. Knowles, J. R. *Science (Washington, D.C.)* **1987**, 236, 1252–1258. Napper, A. D.; Benkovic, S. J.; Tramontano, A.; Lerner, R. A. *Science (Washington, D.C.)* **1987**, 237, 1041–1043.

(4) Whitesides, G. M.; Wong, C.-H. *Angew. Chem., Int. Ed. Engl.* **1985**, 24, 617–638. Jones, J. B. *Tetrahedron* **1986**, 42, 3351–3403.

(10) Kaba, R. A.; Lunazzi, L.; Lindsay, D.; Ingold, K. U. *J. Am. Chem. Soc.* **1975**, 97, 6762.

(11) Nelsen, S. F.; Landis, R. T., II. *J. Am. Chem. Soc.* **1973**, 95, 2719.

(12) Lunazzi, L.; Ingold, K. U. *J. Am. Chem. Soc.* **1974**, 96, 5558.

(13) Malatesta, V.; Ingold, K. U. *J. Am. Chem. Soc.* **1973**, 95, 6110.

(14) Smith, P.; Maples, K. R. *J. Magn. Reson.* **1985**, 65, 491.